



# The Role of SARS-CoV-2 in Male Reproduction

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#### **Abstract**

COVID19 is an infectious disease transmitted by the SARS-CoV-2 virus, whose outbreak was declared a pandemic in March 2020. To date, on November 17, 2020, 55,243,538 confirmed cases had been reported worldwide. Epidemiological studies in different countries have shown higher morbidity and mortality in male than in female patients. The relationship between the COVID-19 disease and the renin-angiotensin-aldosterone (RAA) system has also been documented. The SARS-CoV-2 enters cells through a receptor called angiotensin-converting enzyme-2 (ACE2) and a serine protease (TMPRSS2), both widely expressed in the body, including the testes. ACE2 belongs to the RAA system, which is also expressed in the male reproductive system, and its absence causes infertility. Moreover, ADAM17 is a metalloprotease responsible for inflammation and spermatogenesis and is activated by SARS-CoV-ECA2. Knowledge about the consequences of SARS-CoV-2 infection on male reproduction, as well as the possibility of sexual transmission, is still limited. This review summarizes the available evidence to analyze the effect of SARS-CoV-2 infection on male reproduction and its possible sexual transmission. The reproductive consequences caused by COVID-19 are currently unknown. Although most studies have shown the absence of SARS-COV-2 in the semen and prostate secretion, there is evidence of testicular tissue alteration accompanied by inflammatory infiltration in viral orchitis. These results suggest that there may be a deterioration in the testicular function that could lead to infertility. Also, more studies are needed to assess the risk of sexual transmission.

Keywords: SARS-CoV-2, ACE2, TMPRSS2, ADAM17, Infertility

#### Introduction

Numerous pneumonia cases of unknown etiology appeared at the end of December 2019 in Hubei's province in China, where the virus was identified as a severe respiratory syndrome, named as coronavirus-2 (SARS-CoV-2).<sup>1,2</sup> The World Health Organization (WHO) declared the infection caused by SARS-CoV-2, a pandemic, and named the disease COVID-19. Since its discovery, the new pneumonia caused by this virus continues to spread around the world, with a current count of 55,243,538 confirmed cases and 1,330,930 deaths worldwide.<sup>3</sup> The COVID-19 disease is transmitted mainly by a direct contact with the infected person through the droplets expelled when talking, coughing, or sneezing; and through contact with surfaces contaminated with secretions from the respiratory tract of infected people.4 The incubation period is 6-14 days.5 Viral transmission from presymptomatic and asymptomatic individuals has promoted viral shedding

throughout the world.<sup>6,7</sup> Diverse clinical manifestations appear in the population infected by SARS-CoV-2, some of them include the asymptomatic carrier, the acute respiratory disease, and pneumonia; which present common signs and symptoms, such as fever, dry cough, headache, asthenia, myalgia, sore throat, and less common rhinorrhoea, diarrhea, hemoptysis, nausea, or vomiting, and conjunctival congestion. Anosmia and ageusia have been recognized as prevalent early symptoms in both asymptomatic and COVID-19-positive patients.<sup>8-11</sup>

In early 2020, SARS-CoV-2 was identified as a member of the family Coronaviridae.  $^{1,2}$  Coronaviruses, that includes four genera ( $\alpha$ ,  $\beta$ ,  $\gamma$ , and  $\delta$ ) SARS-CoV-2, SARS-CoV, and MERS-CoV belong to the genus  $\beta$ -coronavirus and can cause acute respiratory distress syndrome in humans.  $^{12}$  SARS-CoV-2 is an enveloped virus with a single-stranded RNA genome, a non-segmented

and positive sense, a part of its genome encodes the spike (S), envelope (E), membrane (M), and nucleocapsid (N). The spike protein (S) binds to the receptor on the host cell membrane's surface, the membrane protein that participates in viral assembly and budding, and the nucleocapsid protein, which binds to the genome and participates in assembly and viral lysis. 13,14 SARS-CoV-2 and SARS-CoV enter cells through the interaction of their protein S with the receptor called angiotensin-converting enzyme (ACE2), 15,16 which is highly present in the human body. It can be found in the oral, nasal and nasopharynx mucosa, lungs, stomach, small intestine, colon, skin, lymph nodes, thymus, bone marrow, spleen, liver, kidney, brain, testis, and prostate. 17-19 Due to the presence of ACE2 in male reproduction organs, SARS-CoV-2 infection may have an impact in male reproduction. The review aims to analyze the available information of the SARS-CoV-2 infection and its potential effects on male fertility, and its sexual transmission. For this purpose, Pubmed, Science Direct and MedRxiv databases were employed to identify publications in this regard.

# **Mechanism of Pathogenesis**

The pathogenesis SARS-CoV mechanism involves the expression of the ACE2 receptor, although SARS-CoV2 has a 20-fold greater affinity to receptor. 16 To get into cells, the S proteins of SARS-Cov-2 associate with the ACE2 receptor. Protein S is formed by two subunits that facilitate the viral-host cell union; the S1 domain interacts and binds through the ACE2 receptor binding domain, while the S2 domain participates in the fusion of the virus-host cell membranes. To complete its entry, S2 is cleaved by the transmembrane protease serine 2 (TMPRSS2), contributing to receptor separation and subsequent membrane fusion, thus facilitating SAR-CoV entry SARS-CoV2 through endocytosis. 14,16,20 Also, protein S binding to ACE2 decreases its expression and the loss of the ACE2 expression turn into a severe acute respiratory failure.<sup>21</sup>

ACE2 is a transmembrane metalloproteinase that shows significant homology to the classic ACE isoform. Both isoforms are part of the renin-angiotensinaldosterone system (RAAS), a critical system that regulates the cardiovascular system and glucose homeostasis among others. <sup>22,23</sup> Renin is a proteolytic enzyme secreted by the kidney in response to blood pressure or sodium concentration. Renin cleaves angiotensinogen, generating angiotensinogen I.<sup>24</sup> While ACE catalyzes angiotensin

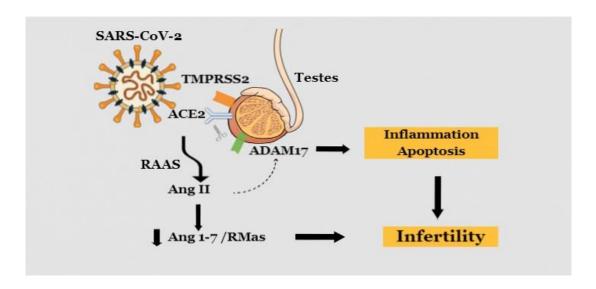
I to angiotensin-II, ACE2 is responsible for the generation of angiotensin 1-9 and 1-7 from angiotensin I and II, respectively.<sup>25</sup> Meanwhile, angiotensin II acts on AT1 receptors producing vasoconstriction, sympathetic activation, inflammation, oxidative stress, and insulin resistance, while angiotensin 1-7 provokes anti-inflammatory, antifibrotic, and diuretic actions through the Mas receptor.<sup>26,27</sup>

# **Role of ACE and TMPRSS2 Isoforms in Testes**

The expression of various RAAS molecules has been identified in testes, epididymis, and spermatozoa in different animal and human species. <sup>25,28–35</sup> Several studies have shown that ACE expression is involved in the processes of spermatogenesis, spermiogenesis, sperm capacitation, as well as in fertilization. <sup>34,36–40</sup> ACE2 is also expressed in spermatogonia, Leydig cells, and Sertoli cells in human testes. <sup>41,42</sup> Angiotensin 1-7 is expressed in Leydig cells and interstitial cells; the Mas receptor is expressed in tubular compartments and the seminiferous epithelium. Likewise, the null expression of ACE shows alterations in male fertility <sup>43</sup>, including the loss of the ability to bind to the zona pellucida of the oocyte <sup>40</sup> and oocyte-sperm fusion. <sup>36,44</sup>

Interestingly, in patients with infertility due to azoospermia, the angiotensin 1-7 and the Mas receptor expression seems to significantly have decreased.<sup>30</sup> Mas receptor deficiency has been shown to be involved in the regulation of spermatogenesis and testicular apoptosis.<sup>29</sup> The participation of ACE2 as a regulator of spermatogenesis and sperm function has been recognized.<sup>39</sup> On the other hand, TMPRSS2 and ACE2 are co-expressed in prostate cells; in particular, the expression of TMPRSS2 in the epithelium of the prostate gland is dependent on androgens and is a component of the seminal fluid proteasome, whose function is associated with the protection of sperm. Furthermore, the expression of TMPRSS2 increases in prostate cancer cells in response to androgens, 45,46 and it has been suggested that the predominance of TMPRSS2 over testosterone contributes to the predominance of COVID-19 in men.47 There are no studies focused on evaluating infection by SARS-CoV2 and the RAAS system that involves the male infertility receptor ACE2. Furthermore, a family of transmembrane metalloproteases known as ADAM has an important role in the processes of fertilization and cellular communication. ADAM17 is a membrane-bound enzyme present in testicular germ cells and is expressed during spermatogenesis and induction of apoptosis by increasing the level of FAS-L on the cell surface.<sup>48</sup> In addition, ADAM17 cleaves and activates various substrates such as receptors of TNF I and II IL-6 participating in the inflammation process and, interestingly, it also cleaves the ACE2 receptor. Likewise, ADAM17 activity is increased by internalization of SARS-CoV2-ACE2 helping viral entry and tissue injury.<sup>49</sup> Also, increased ADAM17 activity has been evidenced with

some comorbidities such as heart failure, chronic lung inflammation, diabetes, and kidney diseases.<sup>50</sup> Therefore, the reduction of ACE2 expression on the cell surface causes an imbalance in the RAAS system, increasing Ang II that induces ADAM17 activity.<sup>26</sup> This metalloprotease is involved in inflammation by cleaving and activating cytokines and cytokine receptors.<sup>51</sup> These results suggest that the processes of inflammation and apoptosis could have an impact on male fertility (Figure 1).



**Figure 1.** Proposal for the involvement of SARS-CoV-2 infection with the male reproductive system. SARS-CoV-2 enters cells through a receptor called angiotensin converting enzyme type 2 (ACE2). The transmembrane protease serine 2 (TMPRSS2), contributes to the fusion of the membrane, facilitating the entry of the virus through endocytosis. Internalization of SARS-CoV2-ACE2 activates a desintegrin and metalloprotease 17 (ADAM17), which participates in inflammation, apoptosis, and cleavage of ACE2. The reduction of ACE2 expression on the cell surface causes an imbalance in the RAAS system, increasing the Ang II that induces ADAM17 activity. The increase in inflammation and apoptosis could have an impact on male fertility.

# SARS-CoV-2 in Male Fertility and Sexual Transmission

A previous study showed that SARS-CoV infection affects the testicles, causing orchitis in humans, showing an alteration of the germ cells with few sperms in the seminiferous tubules accompanied by leukocyte infiltration and abundant IgG in the intestinal tissue.<sup>52</sup> Likewise, another study showed male gonadal dysfunction in 81 patients infected with SARS-CoV-2.<sup>53</sup> This study observed a decrease in the ratio of testosterone/luteinizing hormone, suggesting it as a marker to assess testicular deterioration by SARS-CoV-2.<sup>54</sup> Also, postmortem analysis of testicles from COVID-19 positive patients' showed lymphocytic improvement, decrease in several Leydig cells, lesions in Sertoli cells and seminiferous tubules, with the absence of the virus in the testicular tissue.<sup>54</sup>

Few studies have focused on evaluating the effects of SARS-CoV-2 infection on male reproduction. The existing studies have performed semen analysis in patients recovered from COVID-19 and mild to moderate pneumonia (Table 1). The presence of SARS-CoV-2 in semen is still contradictory. The virus's presence was evidenced in six COVID-19 positive patients (4 in the acute phase and 2 in the recovery phase).55 However, several studies have reported the absence of SARS-CoV-2 in semen and prostate secretion. 56-63 In a SARS-CoV-2 positive patient with moderate symptoms, the virus's presence was not found in the semen eight days after the symptoms began.<sup>62</sup> In another study, patients recovered from COVID-19 reported scrotal discomfort suggesting viral orchitis. However, no virus was seen in the semen a month after patients had been diagnosed

with COVID-19.<sup>57</sup> Similarly, in another study, investigators did not find the presence of SARS-CoV-2 in the semen of 12 patients in the recovery phase with severe symptoms, nor in two patients in the acute phase of COVID-19.<sup>59</sup> Complementing this information, in another study, this virus was not found in recovered patients; most of which had mild complications from COVID-19. Another investigation where semen was collected from patients during the acute infection stage who presented mild discomfort (68%) and moderate pneumonia (32%) as a consequence of SARS-CoV-2, did not find the virus in the semen too. 60 Likewise, there was no evidence of SARS-CoV-2 in the semen of nine patients with an asymptomatic level and an asymptomatic patient.<sup>61</sup> Currently, the absence of SARS-CoV-2 has been shown in the prostate secretion of three COVID-19 positive patients in the nasopharyngeal smear, nor seven previously positive at sampling patients. However, the semen was not evaluated, and the number of patients were limited; therefore, more studies are needed to determinate the prostate alteration.<sup>63</sup>

Approximately, 27 viruses can cause viremia in the human semen.<sup>64</sup> Viruses can cross the blood-testicular barrier formed by Sertoli cells adjacent to the seminiferous tubules' basement membrane, which maintains an essential microenvironment with a unique immunity for testicular function. 64,65 Like SARS-CoV, other viruses such as HIV and Epstein-Barr can cause viral orchitis,<sup>52</sup> germ cell apoptosis, and inflammation. It is worth mentioning that germ cell damage causes infertility,66 in addition to the possibility of contracting sexually transmitted infections.<sup>64</sup> Knowledge about the presence of SARS-CoV-2 in semen is essential due to the clinical and public health implications. There is insufficient evidence to confirm the risk of contagion of SARS-CoV-2 through semen. In addition to the fact that sexual practice is not limited to intercourse, they have recommended abstinence from intercourse, oral or anal sex in SARS-positive patients, because the virus persists in nasal secretions and feces for 10 to 16 days respectively after the absence of COVID-19 symptoms.67

Table 1. SARS-CoV-2 in semen

| Ref | Patients (N) | Appearance of symptoms* (Days) | Sampling**<br>(Days) | Severity of SARS-<br>COV2 infection in<br>patients | Confirmation of COVID19<br>(Gen amplified by PCR) | Presence of viral<br>RNA in semen |
|-----|--------------|--------------------------------|----------------------|--|---|-----------------------------------|
| 55  | 38           | 6-16                           | 2-13                 |  |   | Positive (N=6)                    |
| 62  | 1            | 15                             | 8                    | Mild   | EyS   | Negative                          |
| 57  | 34           | -                              | 31                   | Mild to Moderate                                   | ORF1ab y N  | Negative                          |
| 58  | 18           | -                              | 43.5-47              | Mild (14)***<br>Moderate (4)***                    | -   | Negative                          |
| 56  | 12           | -                              | 14-42                | Mild (92%)***<br>Asymtomatic (8%)***               | <del>-</del>                                      | Negative                          |
| 59  | 23           | -                              | 32                   | Mild (78%)***<br>Moderate (22%) ***                | N y ORF   | Negative                          |
| 60  | 16           | -                              | 0-7                  | Mild (68.7%)***<br>Moderate (31.3%)***             | -   | Negative                          |
| 61  | 9            | -                              | 7-88                 | Mild (8)***<br>Asymptomatic (1)***                 | R, E y N  | Negative                          |
| 63  | 10           | 11                             | 3-23                 |  | N y ORF   | Negative****                      |

<sup>\*</sup> Semen collection days after the onset of symptoms.

### Conclusion

The repercussions on testicular function and infertility caused by the infection of SARS-CoV and SARS-CoV-2 is considerable because the functional receptor ACE2 and TMPRSS2 are expressed in the male reproductive tract. SARS-CoV generates orchitis in humans, whose virus invades cells through the same pathway. ACE2 is an essential part of SAAR, which is

involved in fertilization. Furthermore, ADAM17 is a SARS-CoV-activated metalloprotease and participates in the regulation of spermatogenesis, inflammation, and proteolytic cleavage of ACE2. The expression of molecules such as ACE2/TMPRSS2/ADAM17 in the male reproductive tract denotes that SARS-CoV2 infection could be involved in spermatogenesis. It is essential to analyze the viral load and the conditions

<sup>\*\*</sup> Days of semen collection after confirmatory diagnosis.

<sup>\*\*\*</sup> Classification of patients according to the severity of the infection.

<sup>\*\*\*\*</sup> Negative in prostatic secretion (N=10; 3 positive; 7 recovering patients).

that make SARS-CoV-2 cause a testicular alteration and affect its hormonal regulation. Therefore, it should not be underestimated that COVID-19 disease could lead to male fertility.

#### **Conflict of Interest**

The authors declare no conflict of interest.

#### References

- Lu R, Zhao X, Li J, Niu P, Yang B, Wu H, et al. Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding. Lancet. 2020;395(10224):565-74. doi:10.1016/S0140-6736(20)30251-8
- Chen L, Liu W, Zhang Q, Xu K, Ye G, Wu W, et al. RNA based mNGS approach identifies a novel human coronavirus from two individual pneumonia cases in 2019 Wuhan outbreak. Emerging Microbes & Infections. 2020;9(1):313-9. doi:10.1080/22221751.2020.1725399
- 3. Johns Hopkins University/Coronavirus Resource Center. World MAP. [Internet]. COVID-19 Dashboard by the Center for Systems Science and Engineering (CSSE) at Johns Hopkins University (JHU). Estados Unidos:Johns Hopkins University; 2020. [cited 2020 Oct 19]. Available from: https://coronavirus.jhu.edu/map.html
- Rahman HS, Aziz MS, Hussein RH, Othman HH, Omer SH, Khalid ES, et al. The transmission modes and sources of COVID-19: A systematic review. International Journal of Surgery Open. 2020;26:125-36. doi:10.1016/j.ijso.2020.08.017
- 5. Backer JA, Klinkenberg D, Wallinga J. Incubation period of 2019 novel coronavirus (2019-nCoV) infections among travellers from Wuhan, China, 20–28 January 2020. Eurosurveillance. 2020;25(5):2000062. doi:10.2807/1560-791 7.ES.2020.25.5.2000062
- Wei WE, Li Z, Chiew CJ, Yong SE, Toh MP, Lee VJ. Presymptomatic transmission of SARS-CoV-2—Singapore, january 23–march 16, 2020. Morbidity and Mortality Weekly Report. 2020;69(14):411-15. doi:10.15585/mmwr.mm6914e1
- 7. Bai Y, Yao L, Wei T, Tian F, Jin DY, Chen L, et al. Presumed asymptomatic carrier transmission of COVID-19. Jama. 2020;323(14):1406-7. doi:10.1001/jama.2020.2565
- 8. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet. 2020;395(10223):497-506. doi:10.1016/S0140-6736(20)30183-5
- 9. Zhang JJ, Dong X, Cao YY, Yuan YD, Yang YB, Yan YQ, et al. Clinical characteristics of 140 patients infected with SARS-CoV-2 in Wuhan, China. Allergy. 2020;75(7):1730-41. doi:10. 1111/all.14238
- Ge H, Wang X, Yuan X, Xiao G, Wang C, Deng T, et al. The epidemiology and clinical information about COVID-19. European Journal of Clinical Microbiology & Infectious Diseases. 2020;39(6):1011-9. doi:10.1007/s10096-020-03874-z
- Lechien JR, Chiesa-Estomba CM, De Siati DR, Horoi M, Le Bon SD, Rodriguez A, et al. Olfactory and gustatory dysfunctions as a clinical presentation of mild-to-moderate forms of the coronavirus disease (COVID-19): a multicenter European study. European Archives of Oto-Rhino-Laryngology. 2020;277(8):2251-61. doi:10.1007/s00405-020-05965-1
- 12. Paules Cl, Marston HD, Fauci AS. Coronavirus infections—more than just the common cold. Jama. 2020;323(8):707-8. doi:10.1001/jama.2020.0757
- 13. Chen Y, Liu Q, Guo D. Emerging coronaviruses: genome structure, replication, and pathogenesis. Journal of Medical Virology. 2020;92(4):418-23. doi:10.1002/jmv.25681
- 14. Malik YA. Properties of coronavirus and SARS-CoV-2. The Malaysian Journal of Pathology. 2020;42(1):3-11.
- Lukassen S, Chua RL, Trefzer T, Kahn NC, Schneider MA, Muley T, et al. SARS-CoV-2 receptor ACE 2 and TMPRSS 2 are primarily expressed in bronchial transient secretory cells. The EMBO Journal. 2020;39(10):e105114. doi:10.15252/embj.20 105114
- Hoffmann M, Kleine-Weber H, Schroeder S, Kruger N, Herrler T, Erichsen S, et al. SARS-CoV-2 cell entry depends on ACE2

- and TMPRSS2 and is blocked by a clinically proven protease inhibitor. Cell. 2020;181(2):271-80. doi:10.1016/j.cell.2020.0 2.052
- Li MY, Li L, Zhang Y, Wang XS. Expression of the SARS-CoV-2 cell receptor gene ACE2 in a wide variety of human tissues. Infectious Diseases of Poverty. 2020;9(1):45. doi:10.1186/s40249-020-0 0662-x
- 18. Hamming I, Timens W, Bulthuis ML, Lely AT, Navis GV, van Goor H. Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis. The Journal of Pathology: A Journal of the Pathological Society of Great Britain and Ireland. 2004;203(2):631-7. doi:10.1002/path.1570
- Song H, Seddighzadeh B, Cooperberg MR, Huang FW. Expression of ACE2, the SARS-CoV-2 receptor, and TMPRSS2 in prostate epithelial cells. European Urology. 2020;78(2):296-8. doi:10.1016/j.eururo.2020.04.065
- Matsuyama S, Nagata N, Shirato K, Kawase M, Takeda M, Taguchi F. Efficient activation of the severe acute respiratory syndrome coronavirus spike protein by the transmembrane protease TMPRSS2. Journal of Virology. 2010;84(24):12658-64. doi:10.1128/JVI.01542-10
- 21. Kuba K, Imai Y, Rao S, Gao H, Guo F, Guan B, et al. A crucial role of angiotensin converting enzyme 2 (ACE2) in SARS coronavirus–induced lung injury. Nature Medicine. 2005;11 (8):875-9. doi:10.1038/nm1267
- 22. Luther JM, Brown NJ. The renin–angiotensin–aldosterone system and glucose homeostasis. Trends in Pharmacological Sciences. 2011;32(12):734-9. doi:10.1016/j.tips.2011.07.006
- 23. Forrester SJ, Booz GW, Sigmund CD, Coffman TM, Kawai T, Rizzo V, et al. Angiotensin II signal transduction: an update on mechanisms of physiology and pathophysiology. Physiological Reviews. 2018;98(3):1627-738. doi:10.1152/physrev.00038.2
- 24. Ames MK, Atkins CE, Pitt B. The renin-angiotensin-aldosterone system and its suppression. Journal of Veterinary Internal Medicine. 2019;33(2):363-82. doi:10.1111/jvim.15454
- 25. Donoghue M, Hsieh F, Baronas E, Godbout K, Gosselin M, Stagliano N, et al. A novel angiotensin-converting enzymerelated carboxypeptidase (ACE2) converts angiotensin I to angiotensin 1-9. Circulation Research. 2000;87(5):e1-9. doi:10.1161/01.RES.87.5.e1
- Patel VB, Clarke N, Wang Z, Fan D, Parajuli N, Basu R, et al. Angiotensin II induced proteolytic cleavage of myocardial ACE2 is mediated by TACE/ADAM-17: a positive feedback mechanism in the RAS. Journal of Molecular and Cellular Cardiology. 2014;66:167-76. doi:10.1016/j.yjmcc.2013.11.017
- 27. Santos RÁ, Sampaio WO, Alzamora ÁC, Motta-Santos D, Alenina N, Bader M, et al. The ACE2/angiotensin-(1–7)/MAS axis of the renin-angiotensin system: focus on angiotensin-(1–7). Physiological Reviews. 2018;98(1):505-53. doi:10.1152/physrev.00023.2016
- 28. Al-Maghrebi M, Renno WM. The tACE/angiotensin (1–7)/mas axis protects against testicular ischemia reperfusion injury. Urology. 2016;94:312-e1. doi:10.1016/j.urology.2016.04.021
- 29. Leal MC, Pinheiro SV, Ferreira AJ, Santos RA, Bordoni LS, Alenina N, et al. The role of angiotensin-(1–7) receptor Mas in spermatogenesis in mice and rats. Journal of Anatomy. 2009;214(5):736-43. doi:10.1111/j.1469-7580.2009.01058.x
- 30. Reis AB, Arabjo FC, Pereira VM, Dos Reis AM, Santos RA, Reis FM. Angiotensin (1–7) and its receptor Mas are expressed in the human testis: implications for male infertility. Journal of Molecular Histology. 2010;41(1):75-80. doi:10.1007/s10735-010-9264-8
- 31. Tipnis SR, Hooper NM, Hyde R, Karran E, Christie G, Turner AJ. A human homolog of angiotensin-converting enzyme: cloning and functional expression as a captopril-insensitive carboxypeptidase. Journal of Biological Chemistry. 2000;275(43):33238-43. doi:10.1074/jbc.M002615200
- 32. Pandey KN, Inagami T. Regulation of renin angiotensins by gonadotropic hormones in cultured murine Leydig tumor cells. Release of angiotensin but not renin. Journal of Biological Chemistry. 1986;261(9):3934-8. doi:10.1016/S0021-9258(17)35604-1
- 33. Leung PS, Sernia C. The renin-angiotensin system and male reproduction: new functions for old hormones. Journal of Molecular Endocrinology. 2003;30(3):263-70. doi:10.1677/jme.0.0300263

- 34. Kuhn FM, Dammshauser I, Neukamm C, Renneberg H, Siems WE, Schill WB, et al. Ultrastructural localization of angiotensin-converting enzyme in ejaculated human spermatozoa. Human Reproduction. 1998;13(3):604-10. doi:10.1093/humrep/13.3.604
- Pascolo L, Zito G, Zupin L, Luppi S, Giolo E, Martinelli M, et al. Renin angiotensin system, COVID-19 and male fertility: any risk for conceiving?. Microorganisms. 2020;8(10):1492. doi:10.3390/microorganisms8101492
   Deguchi E, Tani T, Watanabe H, Yamada S, Kondoh G.
- 36. Deguchi E, Tani T, Watanabe H, Yamada S, Kondoh G. Dipeptidase-inactivated tACE action in vivo: selective inhibition of sperm-zona pellucida binding in the mouse. Biology of Reproduction. 2007;77(5):794-802. doi:10.1095/biolreprod.107.060004
- 37. Langford KG, Zhou Y, Russell LD, Wilcox JN, Bernstein KE. Regulated expression of testis angiotensin-converting enzyme during spermatogenesis in mice. Biology of Reproduction. 1993;48(6):1210-8. doi:10.1095/biolreprod
- 38. Nikolaeva MA, Balyasnikova IV, Alexinskaya MA, Metzger R, Franke FE, Albrecht RF, et al. Testicular isoform of angiotensin I-converting enzyme (ACE, CD143) on the surface of human spermatozoa: revelation and quantification using monoclonal antibodies. American Journal of Reproductive Immunology. 2006;55(1):54-68. doi:10.1111/j.1600-0897.2005.00326.x
- 39. Pan PP, Zhan QT, Le F, Zheng YM, Jin F. Angiotensin-converting enzymes play a dominant role in fertility. International journal of Molecular Sciences. 2013;14(10): 21071-86. doi:10.3390/ijms141021071
- 40. Yamaguchi R, Yamagata K, Ikawa M, Moss SB, Okabe M. Aberrant distribution of ADAM3 in sperm from both angiotensin-converting enzyme (Ace)-and calmegin (Clgn)-deficient mice. Biology of Reproduction. 2006;75(5):760-6. doi:10.1095/biolreprod.106.052977
- 41. Douglas GC, O'Bryan MK, Hedger MP, Lee DK, Yarski MA, Smith AI, et al. The novel angiotensin-converting enzyme (ACE) homolog, ACE2, is selectively expressed by adult Leydig cells of the testis. Endocrinology. 2004;145(10):4703-11. doi:10.1210/en.2004-0443
- 42. Wang Z, Xu X. scRNA-seq profiling of human testes reveals the presence of the ACE2 receptor, a target for SARS-CoV-2 infection in spermatogonia, Leydig and Sertoli cells. Cells. 2020;9(4):920. doi:10.3390/cells9040920
- 43. Kessler SP, Rowe TM, Gomos JB, Kessler PM, Sen GC. Physiological non-equivalence of the two isoforms of angiotensin-converting enzyme. Journal of Biological Chemistry. 2000;275(34):26259-64. doi:10.1074/jbc.M004006200
- 44. Li LJ, Zhang FB, Liu SY, Tian YH, Le F, Wang LY, et al. Human sperm devoid of germinal angiotensin-converting enzyme is responsible for total fertilization failure and lower fertilization rates by conventional in vitro fertilization. Biology of Reproduction. 2014;90(6):125. doi:10.1095/biolreprod.113.1 14827
- 45. Chen YW, Lee MS, Lucht A, Chou FP, Huang W, Havighurst TC, et al. TMPRSS2, a serine protease expressed in the prostate on the apical surface of luminal epithelial cells and released into semen in prostasomes, is misregulated in prostate cancer cells. The American Journal of Pathology. 2010;176(6):2986-96. doi:10.2353/ajpath.2010.090665
- 46. Lucas JM, True L, Hawley S, Matsumura M, Morrissey C, Vessella R, et al. The androgen-regulated type II serine protease TMPRSS2 is differentially expressed and mislocalized in prostate adenocarcinoma. The Journal of Pathology: A Journal of the Pathological Society of Great Britain and Ireland. 2008;215(2):118-25. doi:10.1002/path.2330
- 47. Stopsack KH, Mucci LA, Antonarakis ES, Nelson PS, Kantoff PW. TMPRSS2 and COVID-19: serendipity or opportunity for intervention?. Cancer Discovery. 2020;10(6):779-82. doi:10.1158/2159-8290.CD-20-0451
- 48. Lizama C, Rojas-Benhtez D, Antonelli M, Ludwig A, Bustamante-Marhn X, Brouwer-Visser J, et al. TACE/ADAM17 is involved in germ cell apoptosis during rat spermatogenesis. Reproduction. 2010;140(2):305-17. doi:10.1530/REP-10-0104
- 49. Haga S, Yamamoto N, Nakai-Murakami C, Osawa Y, Tokunaga K, Sata T, et al. Modulation of TNF-α-converting enzyme by the spike protein of SARS-CoV and ACE2 induces TNF-α production and facilitates viral entry. Proceedings of the

- National Academy of Sciences. 2008;105(22):7809-14. doi:10.1073/pnas.0711241105
- 50. Zipeto D, Palmeira JD, Argacaraz GA, Argacaraz ER. ACE2/ADAM17/TMPRSS2 interplay may be the main risk factor for COVID-19. Frontiers in Immunology. 2020;11: 576745. doi:10.3389/fimmu.2020.576745
- Dustorhuft S, Lokau J, Garbers C. The metalloprotease ADAM17 in inflammation and cancer. Pathology-Research and Practice. 2019;215(6):152410. doi:10.1016/j.prp.2019.04.002
- 52. Xu J, Qi L, Chi X, Yang J, Wei X, Gong E, et al. Orchitis: a complication of severe acute respiratory syndrome (SARS). Biology of Reproduction. 2006;74(2):410-6. doi:10.1095/biolreprod.105.044776
- Ma L, Xie W, Li D, Shi L, Mao Y, Xiong Y, et al. Effect of SARS-CoV-2 infection upon male gonadal function: a single center-based study. MedRxiv. 2020. doi:10.1101/2020.03.21.2003 7267
- 54. Yang Z, Shi J, He Z, Lu Y, Xu Q, Ye C, et al. Predictors for imaging progression on chest CT from coronavirus disease 2019 (COVID-19) patients. Aging. 2020;12(7):6037-48. doi:10.18632/aging.102999
  55. Li D, Jin M, Bao P, Zhao W, Zhang S. Clinical characteristics
- 55. Li D, Jin M, Bao P, Zhao W, Zhang S. Clinical characteristics and results of semen tests among men with coronavirus disease 2019. JAMA Network Open. 2020;3(5):e208292. doi:10.1001/jamanetworkopen.2020.8292
- 56. Song C, Wang Y, Li W, Hu B, Chen G, Xia P, et al. Detection of 2019 novel coronavirus in semen and testicular biopsy specimen of COVID-19 patients. MedRxiv. 2020. doi:10.1101/2020.03.31.20042333
- 57. Pan F, Xiao X, Guo J, Song Y, Li H, Patel DP, et al. No evidence of severe acute respiratory syndrome—coronavirus 2 in semen of males recovering from coronavirus disease 2019. Fertility and Sterility. 2020;113(6):1135-9. doi:10.1016/j.fertnster t.2020.04.024
- 58. Holtmann N, Edimiris P, Andree M, Doehmen C, Baston-Buest D, Adams O, et al. Assessment of SARS-CoV-2 in human semen—a cohort study. Fertility and Sterility. 2020;114(2):233-8. doi:10.1016/j.fertnstert.2020.05.028
- Guo L, Zhao S, Li W, Wang Y, Li L, Jiang S, et al. Absence of SARS-CoV-2 in semen of a COVID-19 patient cohort. Andrology. 2021;9(1):42-7. doi:10.1111/andr.12848
- 60. Kayaaslan B, Korukluoglu G, Hasanoglu I, Kalem AK, Eser F, Akinci E, et al. Investigation of SARS-CoV-2 in semen of patients in the acute stage of COVID-19 infection. Urologia Internationalis. 2020;104(9-10):678-83. doi:10.1159/0005105
- 61. Pavone C, Giammanco GM, Baiamonte D, Pinelli M, Bonura C, Montalbano M, et al. Italian males recovering from mild COVID-19 show no evidence of SARS-CoV-2 in semen despite prolonged nasopharyngeal swab positivity. International Journal of Impotence Research. 2020;32(5):560-2. doi:10.1038/s41443-020-00344-0
- 62. Paoli D, Pallotti F, Colangelo S, Basilico F, Mazzuti L, Turriziani O, et al. Study of SARS-CoV-2 in semen and urine samples of a volunteer with positive naso-pharyngeal swab. Journal of Endocrinological Investigation. 2020;43(12):1819-22. doi:10.1007/s40618-020-01261-1
- 63. Zhang S, Wang X, Zhang H, Xu A, Fei G, Jiang X, et al. The absence of coronavirus in expressed prostatic secretion in COVID-19 patients in Wuhan city. Reproductive Toxicology. 2020;96:90-4. doi:10.1016/j.reprotox.2020.06.006
- 64. Salam AP, Horby PW. The breadth of viruses in human semen. Emerging Infectious Diseases. 2017;23(11):1922-24. doi:10.3201/eid2311.171049
- 65. Zhao S, Zhu W, Xue S, Han D. Testicular defense systems: immune privilege and innate immunity. Cellular & Molecular Immunology. 2014;11(5):428-37. doi:10.1038/cmi.2014.38
- Olaniyan OT, Dare A, Okotie GE, Adetunji CO, Ibitoye BO, Bamidele OJ, et al. Testis and blood-testis barrier in Covid-19 infestation: role of angiotensin-converting enzyme 2 in male infertility. Journal of Basic and Clinical Physiology and Pharmacology. 2020;31(6):20200156. doi:10.1515/jbcpp-202 0.0156
- 67. Cabello F, Sanchez F, Farre JM, Montejo AL. Consensus on recommendations for safe sexual activity during the COVID-19 coronavirus pandemic. Journal of Clinical Medicine. 2020; 9(7):2297. doi:10.3390/jcm9072297